Package: imsig (via r-universe)

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Type Package

Title Immune Cell Gene Signatures for Profiling the Microenvironment
of Solid Tumours

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Description Estimate the relative abundance of tissue-infiltrating
immune subpopulations abundances using gene expression data.

License GPL-3

URL <https://github.com/ajitjohnson/imsig/>

BugReports <https://github.com/ajitjohnson/imsig/issues>

Encoding UTF-8

LazyData true

Imports HiClimR (>= 1.2), RColorBrewer (>= 1.1), igraph (>= 1.2),
ggplot2 (>= 2.2), gridExtra (>= 2.3), survival (>= 2.4)

RoxygenNote 7.1.1

Suggests testthat

Repository https://ajitjohnson.r-universe.dev

RemoteUrl https://github.com/ajitjohnson/imsig

RemoteRef HEAD

RemoteSha 929f5ce9df576b9821c89bee453f16eb30049e05

Contents

corr_matrix   *Correlation matrix*

Description

Creates a correlation matrix of ImSig signature genes.

Usage

```
corr_matrix(exp, r)
```

Arguments

exp   Dataframe of transcriptomic data (natural scale) containing genes as rows and
             samples as columns. Note: Gene names should be set as row names and du-
             plicates are not allowed. Missing values are not allowed within the expression
             matrix. Check example- head(example_data): example_data.

r   Use a value between 0 and 1. Default is 0.6. This is a user defined correlation
             cut-off to perform feature selection (feature_select). Feature selection aids
             to enrich the prediction of relative abundance of immune cells by filtering off
             poorly correlated ImSig genes. To get an idea of what cut-off to use check the
             results of (gene_stat) and choose a cut-off that displays high median correla-
             tion and maintains a high proportion of genes after feature selection.

Value

Gene-gene correlation matrix of ImSig genes.

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---

example_cli   *Example clinical data file for survival analysis with ImSig*

Description

An example clinical data file. Minimum required informations are the sample name (same as that of the expression matrix), event (dead or alive) and time to event (days, months or years).

Usage

```
example_cli
```

Format

dataframe

---

example_data   *Example transcriptomics data*

Description

Example expression data matrix. The data is preffered to be in natural scale with genes as rows and samples as columns.Note: Gene names should be set as row names and duplicates are not allowed. Missing values are not allowed within the expression matrix. Check example- head(example_data)

Usage

```
example_data
```

Format

dataframe

---

feature_select   *Feature selection of signature genes*

Description

ImSig genes were designed to be co-expressed in tissue transcriptomic data. However, depending on the dataset some of the genes may not co-express with the dominant module. In order to remove such deviant genes, a feature selection can be carried out based on correlation. This function removes genes that exhibit a poor correlation (less than the defined r value) with the dominant ImSig module. This step of feature selection is recommended to enrich the prediction of relative abundance of immune cells.

Usage

```
feature_select(exp, r = 0.6)
```

Arguments

exp   Dataframe of transcriptomic data (natural scale) containing genes as rows and samples as columns. Note: Gene names should be set as row names and duplicates are not allowed. Missing values are not allowed within the expression matrix. Check example- head(example_data): example_data.

r   Use a value between 0 and 1. Default is 0.6. This is a user defined correlation cut-off to perform feature selection. To get an idea of what cut-off to use check the results of (gene_stat) and choose a cut-off that displays high median correlation and maintains a high proportion of genes after feature selection.

Value

Returns a list of 'feature selected' genes based on the set r value.

Examples

```
feature_select (exp = example_data, r = 0.7)
```

---

gene_stat   *General stastitics of ImSig analysis*

Description

[Total genes in ImSig]: The total number of genes in ImSig list. [No. of ImSig genes in user dataset]: The number of ImSig genes found in user's dataset. Like all signatures, ImSig works best when this overlap is high, preferably over 75

Usage

```
gene_stat(exp, r = 0.6)
```

Arguments

exp	Dataframe of transcriptomic data (natural scale) containing genes as rows and samples as columns. Note: Gene names should be set as row names and duplicates are not allowed. Missing values are not allowed within the expression matrix. Check example- head(example_data): example_data.
r	Use a value between 0 and 1. Default is 0.6. This is a user defined correlation cut-off to perform feature selection (feature_select). Feature selection aids to enrich the prediction of relative abundance of immune cells by filtering off poorly correlated ImSig genes. To get an idea of what cut-off to use check the results of (gene_stat) and choose a cut-off that displays high median correlation and maintains a high proportion of genes after feature selection.

Value

Dataframe of general statistics of ImSig analysis.

See Also

feature_select

Examples

```
gene_stat (exp = example_data, r = 0.7)
```

---

imsig	*Estimate the relative abundance of tissue-infiltrating immune subpopulations abundances using gene expression data*

Description

Estimates the relative abundance of immune cells across patients/samples.

Usage

```
imsig(exp, r = 0.6, sort = TRUE, sort_by = "T cells")
```

Arguments

exp	Dataframe of transcriptomic data (natural scale) containing genes as rows and samples as columns. Note: Gene names should be set as row names and duplicates are not allowed. Missing values are not allowed within the expression matrix. Check example- head(example_data): example_data.

r	Use a value between 0 and 1. Default is 0.6. This is a user defined correlation cut-off to perform feature selection ([feature_select](#)). Feature selection aids to enrich the prediction of relative abundance of immune cells by filtering off poorly correlated ImSig genes. To get an idea of what cut-off to use check the results of ([gene_stat](#)) and choose a cut-off that displays high median correlation and maintains a high proportion of genes after feature selection.
sort	Sort the samples based on abundance of a particular cell type. 'Set sort = FALSE' if you wish not to apply sorting. By default the function sorts by abundance of T cells. The cell type of interest for sorting can be controlled by the 'sort_by' parameter.
sort_by	Can be used to sort the samples by predicted abundance of a particular cell type. All other cell types follow this sorting. By default it is sorted by 'T cells'

Value

Relative abundance of immune cells across samples. Returns a dataframe.

See Also

[feature_select](#), [example_data](#)

Examples

```
cell_abundance = imsig (exp = example_data, r = 0.7, sort=TRUE, sort_by='T cells')
head(cell_abundance)
```

---

imsig_survival	*Survival analysis based on relative abundance of immune infiltration estimated by ImSig*

Description

Patients are split into two groups based on their immune cell abundance (median aundance value) and a regular survival analyis is carried out.

Usage

```
imsig_survival(exp, cli, time = "time", status = "status", r = 0.6)
```

Arguments

exp	Dataframe of transcriptomic data (natural scale) containing genes as rows and samples as columns. Note: Gene names should be set as row names and duplicates are not allowed. Missing values are not allowed within the expression matrix. Check example- head(example_data): [example_data](#).
cli	Clinical metadata containting the event data (dead or alive) and time to event data. Samples names should be in rownames and same as that in the expression file. Check head() of [example_cli](#) for an example clinical data.

time   Column name of time-to-event parameter.

status   Column name of event (dead or alive) parameter.

r   Use a value between 0 and 1. Default is 0.6. This is a user defined correlation
                    cut-off to perform feature selection ([feature_select](#)). Feature selection aids
                    to enrich the prediction of relative abundance of immune cells by filtering off
                    poorly correlated ImSig genes. To get an idea of what cut-off to use check the
                    results of ([gene_stat](#)) and choose a cut-off that displays high median correla-
                    tion and maintains a high proportion of genes after feature selection.

Value

Hazard Ratio

See Also

[feature_select](#), [example_data](#), [example_cli](#)

Examples

```
survival = imsig_survival (exp = example_data, cli = example_cli)
head(survival)
```

---

plot_abundance   *Plot relative abundance of immune cells*

Description

Barplots of relative abundance of immune cells across samples.The order of the samples are the
same as that of [imsig](#).

Usage

```
plot_abundance(exp, r = 0.6)
```

Arguments

exp   Dataframe of transcriptomic data (natural scale) containing genes as rows and
                    samples as columns. Note: Gene names should be set as row names and du-
                    plicates are not allowed. Missing values are not allowed within the expression
                    matrix. Check example- head(example_data): [example_data](#).

r   Use a value between 0 and 1. Default is 0.6. This is a user defined correlation
                    cut-off to perform feature selection ([feature_select](#)). Feature selection aids
                    to enrich the prediction of relative abundance of immune cells by filtering off
                    poorly correlated ImSig genes. To get an idea of what cut-off to use check the
                    results of ([gene_stat](#)) and choose a cut-off that displays high median correla-
                    tion and maintains a high proportion of genes after feature selection.

Value

ggplot

See Also

[feature_select](), [example_data]()

Examples

```
plot_abundance (exp = example_data, r = 0.7)
```

---

plot_network   *Network graph of ImSig genes*

Description

A Network visualization displays undirected graph structures and highlights the relationships between entities. The nodes are ImSig genes and the edges represent the correlation between them. The nodes are coloured based on cell type. Try using a correlation cut-off of '0' to get a complete picture.

Usage

```
plot_network(
  exp,
  r = 0.6,
  pt.cex = 2,
  cex = 1,
  inset = 0,
  x.intersp = 2,
  vertex.size = 3,
  vertex.label = NA,
  layout = layout_with_fr
)
```

Arguments

exp   Dataframe of transcriptomic data (natural scale) containing genes as rows and
             samples as columns. Note: Gene names should be set as row names and du-
             plicates are not allowed. Missing values are not allowed within the expression
             matrix. Check example- head(example_data): [example_data]().

r   Use a value between 0 and 1. Default is 0.6. This is a user defined correlation
             cut-off to perform feature selection ([feature_select]()). Feature selection aids
             to enrich the prediction of relative abundance of immune cells by filtering off
             poorly correlated ImSig genes. To get an idea of what cut-off to use check the
             results of ([gene_stat]()) and choose a cut-off that displays high median correla-
             tion and maintains a high proportion of genes after feature selection.

pt.cex	expansion factor(s) for the points.
cex	character expansion factor relative to current par("cex"). Used for text, and provides the default for pt.cex.
inset	inset distance(s) from the margins as a fraction of the plot region when legend is placed by keyword.
x.intersp	character interspacing factor for horizontal (x) spacing.
vertex.size	Node size of network graph
vertex.label	Add gene names to the network graph. Default set to NA.
layout	Layout algorithm to be used for building network. Default set to force-directed layout algorithm by Fruchterman and Reingold. Read documentation of 'igraph' for other available algorithms.

Value

Network graph

See Also

[feature_select](#)

Examples

```
plot_network (exp = example_data, r = 0.7)
```

---

plot_survival	*Forest plot of survial analysis by ImSig*

Description

Patients are split into two groups based on their immune cell abundance (median aundance value) and a regular survival analyis is carried out. Raw values can be obtained from [imsig_survival](#).

Usage

```
plot_survival(exp, cli, time = "time", status = "status", r = 0.6)
```

Arguments

exp	Dataframe of transcriptomic data (natural scale) containing genes as rows and samples as columns. Note: Gene names should be set as row names and duplicates are not allowed. Missing values are not allowed within the expression matrix. Check example- head(example_data): [example_data](#).
cli	Clinical metadata containting the event data (dead or alive) and time to event data. Samples names should be in rownames and same as that in the expression file. Check head() of [example_cli](#) for an example clinical data.

time	Column name of time-to-event parameter.
status	Column name of event (dead or alive) parameter.
r	Use a value between 0 and 1. Default is 0.6. This is a user defined correlation cut-off to perform feature selection ([feature_select](#)). Feature selection aids to enrich the prediction of relative abundance of immune cells by filtering off poorly correlated ImSig genes. To get an idea of what cut-off to use check the results of ([gene_stat](#)) and choose a cut-off that displays high median correlation and maintains a high proportion of genes after feature selection.

Value

Forest plot

See Also

[feature_select](#), [example_data](#), [example_cli](#)

Examples

```
plot_survival (exp = example_data, r = 0.7, cli = example_cli, time = 'time', status= 'status')
```

---

pp_exp   *Pre-processing expression matrix*

Description

Subsets the user's dataset based on the genes that are common to the users dataset and ImSig.

Usage

```
pp_exp(exp)
```

Arguments

exp	Dataframe of transcriptomic data (natural scale) containing genes as rows and samples as columns. Note: Gene names should be set as row names and duplicates are not allowed. Missing values are not allowed within the expression matrix. Check example- head(example_data): [example_data](#).

Value

Expression dataframe

---

pp_sig   *Pre-processing ImSig file*

Description

Subsets ImSig genes based on the genes that are common to the users dataset and ImSig

Usage

```
pp_sig(exp)
```

Arguments

exp   Dataframe of transcriptomic data (natural scale) containing genes as rows and
               samples as columns. Note: Gene names should be set as row names and du-
               plicates are not allowed. Missing values are not allowed within the expression
               matrix. Check example- head(example_data): example_data.

Value

ImSig dataframe

---

sig   *ImSig genes*

Description

ImSig signature genes and the cell type they represent

Usage

```
sig
```

Format

dataframe

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